

## Hek 293S Transfection

### **The day before transfection**

Culture 293 cells to a density of  $2 \times 10^6$  cells/ml in DMEM-F12 + 10% FBS + 1% anti-anti

### **I. Transfection day {Morning procedure to avoid late night} (Invitrogen)**

- 1.) Count the cells ( $1.5-2 \times 10^6$  cells/ml to make sure they're in log phase)
- 2.) Seed each well of a 6-well plate with 2ml of  $0.6 \times 10^6$  cells/ml to achieve ~70% confluence. Allow the cells to attach for ~15 minutes at 27-28 C {Note: Prepare transfection mixture in waiting time.}
- 3.) Remove the old medium, wash with Opti-MEM medium once
- 4.) Add 2ml of Opti-MEM medium
- 5.) Add combined transfection mixture (~200ul) dropwise onto each well, and incubate at 37C overnight at 5% CO<sub>2</sub>.

### ***Preparation of Transfection mixture for each well***

- 1.) Prepare (a) Lipofectamine 2000: 8ul (Lipofectamine) + 100ul Opti-MEM medium in a microfuge.
- 2.) Prepare (b) DNA: 1ul (1-2.5ug pQCXIP-GOI) + 100ul Opti-MEM in a microfuge.
- 3.) Add preparation (a) and (b) and incubate 5 min at room temperature in the fume hood.

Tube (a) =  $(100 \times 6 = 600) + (8 \times 6 = 48) = 648\text{ul}$

Tube (b) =  $(100 \times 6 = 600) = (1 \times 6 = 6) = 606\text{ul}$

Tube (a) + (b) = ~1200ul

### **Harvesting**

- 1.) After 12-16 hours incubation time, aspirate the transfection mixture. Add DMEM-F12 + 10% FBS + 1% anti-anti to the cells (2ml in each well). No wash step necessary.
- 2.) Add 10mM sodium butyrate and drop the temperature to 30 C
- 3.) Harvest 48 hours later